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TITLE: Breast Cancer Resource for Research and Banking, with Emphasis on Early Tumors and Precursor Lesions

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13. ABSTRACT (Maximum 200) The Breast Cancer Resource for Research and Banking has accrued breast cells or tissues from 1,467 patients during the grant period (3 year grant with 9 month extension without additional funding). The emphasis of this project has been on the collection of microscopic at risk and precursor lesions as imprints/scraps. Additionally, throughout the grant period, all invasive carcinomas with available tissue have been accrued, since most investigators who have requested samples have requested frozen pieces of tumor tissue paired with normal tissue from the same patient, i.e. their interest has been in established carcinomas and not in precursor lesions. During the last year we also filled requests for specimens for microdissection. Over the entire grant period 26 investigator requests for tissue have been filled. At termination of the grant the Breast Cancer Resource was transferred to the auspices of the Resource for Tumor Tissue and Data of the NYU Kaplan Comprehensive Cancer Center.				
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FOREWORD

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____ In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

____ In the conduct of research utilizing recombinant DNA, the investigator(s) adhered to the NIH Guidelines for Research Involving Recombinant DNA Molecules.

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Robert Fenn MD 9/28/98
PI - Signature Date

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INTRODUCTION

Basic, clinical, and translational research on breast cancer in the United States has been stimulated in recent years by increased government and private funding. Translational research, in particular, requires the availability of human breast cancer tissue, as well as breast tissue with "precursor" and "at risk" lesions. At risk lesions are the proliferative and atypical proliferative components of fibrocystic change, and the precursor lesion is carcinoma in situ. These lesions have been defined histologically and their roles in breast carcinogenesis have been validated epidemiologically. This grant was funded in the category of "Infrastructure Enhancement" specifically to make breast cancer tissue, precursor, and at risk lesions available to investigators in the field of human breast carcinogenesis.

METHODS

Clinical cancers, that is invasive carcinomas that have resulted in a palpable mass lesion, were banked in standard fashion as snap-frozen pieces of tissue, together with pieces of non-neoplastic breast tissue from the same patient, and a portion of lymph node when available.

Collecting at risk and precursor lesions of breast cancer, which are almost invariably microscopic, is difficult, firstly because the lesions are so small, and secondly because good medical practice requires that all the tissue excised be subjected to routine histopathologic examination in order to properly classify the lesion.

Accordingly, in years 1 to 3 of the grant we collected at risk and precursor lesions of breast cancer as slide imprints/scrapes prepared from excised breast tissue prior to histopathologic examination. By the end of year two 782 imprint samples had been collected from mammographically detected (non-palpable) lesions. These covered the spectrum of fibrocystic change (non-proliferative, proliferative, and proliferative with atypia), as well as ductal carcinoma in situ and lobular carcinoma in situ. It was disappointing that despite considerable effort to publicize this collection of material, investigators did not request it. The reasons were twofold. Firstly, most basic scientists are unfamiliar with the histopathologically defined at risk and precursor lesions of breast cancer; they are more interested in established cancers (mass lesions). Secondly, the samples are small and comprised of mixtures of cells (of necessity stromal cells and lymphohistiocytic cells are admixed with lesional cells in imprint/aspirate specimens). At about the same time the technique of microdissection was evolving, and provided an alternate method for acquiring such lesions for research purposes. Accordingly in year 3 and in the extension period we have been supplying investigators with material prepared for microdissection. This technique allows one to obtain pure specimens of microscopic precursor and at risk lesions, from either fixed paraffin embedded tissue (for DNA - PCR studies) or from frozen sections (for RNA based studies). Specimens obtained by microdissection are superior to imprints and aspirates inasmuch as the histologic context from which samples are obtained can be documented, and the samples are pure. Because of the aforesaid, in year 3 we turned our efforts away from imprints and toward providing samples for microdissection.

Use of the Resource has been stimulated by the award of 16 pilot projects from developmental funds from the Kaplan Comprehensive Cancer Center's NCI Breast Cancer Program Grant during the 1995-1998 period.

Outside NYU, the Resource has been included in the Breast Cancer Specimen and Data Information System, a collaborative project sponsored by the National Action Plan for Breast Cancer Biologic Resources Banks Working Group and the NCI. The DOD Breast Cancer Research Program "Era of Hope" in Washington D.C. in October/November 1997 provided another forum for publicizing the Resource.

To obtain feedback on the satisfaction of investigators with the material sent to them, two contacts are made with each recipient. The first is to determine the state of material given or shipped and occurs within a day or two of shipping. The second contact is made 6 - 18 months later to determine the level of satisfaction in terms of results obtained. User files are maintained for each recipient of samples. User records are initiated with "Investigator Request" forms (Appendix 1).

To obtain information on the quality and durability of banked tissues and cells, specimens obtained in 1995, 1996 and 1997 have been subjected to a variety of analyses. These analyses were immunohistochemistry, immunofluorescence microscopy, fluorescence in situ hybridization, and RT-PCR. Analyses were done in various laboratories at NYU that have expertise in these assays. Records are maintained on "Evaluation of Banked Material" forms (Appendix 2).

The Resource technician has also culled the NYU departmental records retrospectively so that all patients with mammographically detected lesions from 1991-1994 have been entered into the database. Even though no fresh samples (imprints/aspirates) are available in these cases, the ability to microdissect the archival samples has made them a valuable addition to the Resource.

RESULTS

The numbers of the various types of breast tissue samples that have been banked and entered into our database during each grant year, as well as the cumulative numbers of samples for the entire collection period are shown in Tables 1 to 4. Tables 1 and 2 use the format of previous annual reports. Tables 3 and 4 show data for all four years.

In Table 1 the breakdown is by type of samples available. In Table 2 the breakdown is by type of lesion as defined histopathologically. Total number of samples in Table 1 exceeds total number of cases in Table 2 because some cases (patients) generated more than one sample type.

TABLE 1

BANKED CASES OF BREAST CANCER AND PRECURSOR LESIONS
AT NYU MEDICAL CENTER BY SAMPLE TYPES

	Grant Year #4 <u>12/97 - 8/98</u>	4 Yr. Cumulative <u>12/94 - 8/98</u>
Imprints/scrapes	0	633
Aspirated cells	115	642
Snap frozen tissue fragments*	<u>69</u>	<u>757</u>
TOTAL	184	2,032

*includes 308 paired samples of breast cancers with normal tissue.

TABLE 2

BANKED CASES OF BREAST CANCER AND PRECURSOR LESIONS
AT NYU MEDICAL CENTER BY HISTOPATHOLOGIC DIAGNOSIS

	Grant Year #4 <u>12/97 - 8/98</u>	4 Yr. Cumulative <u>12/94 - 8/98</u>
Invasive ductal carcinoma	67	383
Invasive lobular carcinoma	12	59
Ductal carcinoma in situ*	7	151
Lobular carcinoma in situ*	0	46
Secondary carcinoma, lymph node	27	110
Lymph node without tumor	0	81
Fibrocystic change, non-proliferative	1	190
Fibrocystic change, proliferative**	1	215
Fibrocystic change, proliferative with atypia**	0	80
Other (mostly fibroadenoma)	<u>13</u>	<u>153</u>
TOTAL	128	1,468

*precursor lesion **at risk lesion

Table 5 indicates the number of patients from whom samples were obtained during years 1 and 3 and the extension period, and during the entire grant period. Table 6 indicates the numbers of requests for specimens that have been filled over similar time periods.

TABLE 3

SPECIMENS BY SAMPLE TYPE

	<u>1995</u>	<u>1996</u>	<u>1997</u>	<u>1998</u>	<u>Total</u>
Imprints/TP	367	415	102	0	633
Aspirates	149	219	159	115	642
Tissue	<u>233</u>	<u>199</u>	<u>256</u>	<u>69</u>	<u>757</u>
TOTAL	749	833	517	184	2,032

TABLE 4

SPECIMENS BY HISTOPATHOLOGIC DIAGNOSIS

	<u>1995</u>	<u>1996</u>	<u>1997</u>	<u>1998</u>	<u>Total</u>
Invasive ductal carcinoma	118	112	86	67	383
Invasive lobular carcinoma	16	20	11	12	59
In situ ductal	55	50	39	7	151
In situ lobular	11	25	10	0	46
Secondary carcinoma	25	23	35	27	110
FCD - proliferative	86	92	90	1	269
FCD - non-proliferative	71	107	11	1	190
Other	<u>48</u>	<u>104</u>	<u>97</u>	<u>13</u>	<u>262</u>
TOTAL	430	533	379	128	1,470

TABLE 5

NUMBER OF PATIENTS WITH BANKED SAMPLES

<u>Grant Year #1</u>	<u>Grant Year #2</u>	<u>Grant Year #3</u>	<u>Grant Year #4</u>	<u>4 Yr. Cumulative</u>
<u>12/94 - 11/95</u>	<u>12/95 - 11/96</u>	<u>12/96 - 11/97</u>	<u>12/97 - 8/98</u>	<u>12/94 - 8/98</u>
430	537	365	135	1,467

TABLE 6

REQUESTS FOR SPECIMENS FILLED

	Grant Yr. #1	Grant Yr. #2	Grant Yr. #3	Grant Yr. #4	4 Yr. Cumulative
	<u>12/94 - 11/95</u>	<u>12/95 - 11/96</u>	<u>12/96 - 11/97</u>	<u>12/97 - 8/98</u>	<u>12/94 - 8/98</u>
Imprints/scrapes	1	1	0	0	2
Frozen tissue	2	4	9	4	19
Tissue for microdissection	<u>0</u>	<u>0</u>	<u>2</u>	<u>3</u>	<u>5</u>
TOTAL	3	5	11	7	26

As shown in Table 3, we reduced the numbers of imprint/scrape samples collected in years 3 and 4 of the grant. These represent samples of microscopic lesions, mainly in situ carcinoma and proliferative fibrocystic changes. The reason for this reduced collection is twofold. Firstly, we now have a large collection of these lesions and requests for such samples have been very low. Secondly, the technique of microdissection has been gaining increasing favor as an alternative method for obtaining samples of microscopic lesions. Current amplification techniques allow the analysis of cells from a single microdissected duct or lobule of breast tissue. Microdissection can be done on frozen or on fixed, paraffin embedded tissue. Furthermore, the purity of specimens can be monitored by examination of sections before and after the microdissection is done. The success of this technique may be the reason for the underutilization of our imprint/scrape samples. Prior to the use of microdissection, scrapes/imprints represented the only means for obtaining precancerous and microscopic breast lesions for research purposes. The disadvantages of imprint/scrapes as compared to microdissection relates to the fact that imprint/scrape samples represent mixtures of cells, albeit the lesional cells predominate. In both instances the samples are small, but investigators prefer to use samples of known and verifiable purity.

There have been several opportunities for publicizing the Resource at NYU. It has been written up three times in the Kaplan Comprehensive Cancer Center newsletter, "LAB NOTES". The principal investigator has lectured on the Resource to the Kaplan Comprehensive Cancer Center Core Grant Working Group and at the NYU Breast Cancer Research Program (BCRP). She is also a major participant at the NYU monthly clinical multidisciplinary breast cancer conferences and a member of the Executive Committee of the NYU Breast Center, both of which provide forums for continually updating colleagues on the size of the Resource and the spectrum of available material. Additionally, the Kaplan Comprehensive Cancer Center Breast Cancer Research Program Grant has funded pilot projects for translational research from 1995-1998 generating intramural users (Appendix 3).

Our Internet listing through the National Action Plan has generated 6 outside users of the Resource, one in 1996, three in 1997, and two in 1998.

Based on investigator feedback, our efforts in filling requests for specimens and determining investigator satisfaction with specimens has produced results ranging from good to

excellent. All investigators have been very satisfied with the state in which they have received specimens shipped or delivered to them. Feedback from 1995, 1996, and early 1997 recipients indicates that the material was suitable for the research techniques that they used. An example of such feedback and publications referring to the Resource and its funding source are shown in Appendix 1.

Several slide-based techniques performed in the principal investigator's department and elsewhere in the Medical Center produced good results of immunohistochemistry (Appendix 2) and immunofluorescence microscopy on archived samples. Fluorescent in situ hybridization (Appendix 2) results have been excellent on 1997 samples, and good on 1995 and 1996 samples.

At termination of the grant the Breast Tissue Resource is being transferred to the auspices of the Resource for Tumor Tissue and Data of the Kaplan Comprehensive Cancer Center. Thus, the Resource technician, materials, and database will remain available. Collection of samples will continue and the materials collected will remain available to investigators.

CONCLUSIONS

The Resource has acquired 2,032 specimens from 1,467 patients.

Requests for snap frozen samples of established breast cancers, matched with normal tissue from the same patient are the most frequent requests received.

Sample preservation is good.

We have met investigator's needs in all instances.

The Resource has provided the principal investigator with outstanding opportunities for ongoing collaboration in various aspects of breast cancer research (1-13).

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LIST OF PERSONNEL RECEIVING PAY FROM THIS EFFORT

Helen Feiner, M.D.
 Jaishree Jagirdar, M.D.
 Ms. Yara Delgado

Feiner, Helen, D.
DAMDM17-94-J-4177

best available copy

APPENDIX 1



Request for Tissue/Cells
NYU Breast Cancer Resource
Director: Helen D. Feiner, M.D.
560 First Ave. NY, NY 10016
Tel (212) 263-8826
Fax (212) 263-7916

Name:	Dr. CHARLES CARMECI/ D. THOMPSON Ph.D
Title:	MD/Ph.D
Address:	STANFORD UNIVERSITY. DEPTO OF SURGERY MSLS P229. 1201 WELCH RD STANFORD, CA 94305
Phone:	(415) 725-1671 (415) 498-5510
Fax:	(415) 725-8762
e-mail:	--
Grant Support:	YES
Material Requested:	BREAST TISSUE IN VIAL CONFIRMED ER+ AND ER- Date Shipped: 8-5-96 4-21-97 Date Received: NEXT DAY State of Specimen on receipt: GOOD
Brief Summary of intended use: (Use additional page if necessary)	TO IDENTIFY AND CHARACTERIZE GENES THAT ARE COORDINATELY EXPRESSED WITH ER AND DETERMINE THEIR INFLUENCE ON BREAST CANCER PROTOTYPE.

Charles Carmeci, MD
Stanford University
Dept of Surgical Oncology
MSLS P229
1201 Welch Rd
Stanford, CA 94305

8/16/96

CL (415) 725-1671

Helen Feiner, MD
Dept of Surgical Pathology
NYU Medical Center
560 First Avenue
New York, NY 10016

Dear Dr. Feiner,

Thank you for the primary breast cancer specimens. They arrived in excellent condition.

We have recently isolated and partially characterized several genes from breast cancer cell lines which are coordinately expressed with the gene for estrogen receptor. We feel that this set of genes plays a critical role in determining the differing phenotypes between ER positive and ER negative carcinomas. Using Northern blots from the samples which you have provided, we aim to determine the expression of these genes in primary tumors. The NIH has provided funding for this project (Grant #: NIH/NRSA#1F32CA69715-01A1 PI: Ronald Weigel, MD, PhD).

Thank you for providing such a valuable resource.

Charles Carmeci
Charles Carmeci, MD

By phone from Dr Carmeci 5/5/97
- yield of 20-30 ng total RNA
- Not great for Northern
- RT PCR yield good of all
ER+ samples and most
ER- samples. Low
yield from normal breast tissue
H

NEW YORK UNIVERSITY MEDICAL CENTER
Anatomic Pathology, Room 461
560 First Avenue
New York, N.Y. 10016

F A X

Date: 1/21/97

Number of pages including cover sheet: 1

TO: DR CHARLES CARMECI FROM: DR HELEN FEINER

Phone: _____

Phone: (212) 263-5470

Fax: 415 723-8762

Fax: (212) 263-7916

As discussed, information to add to
your publication(s) :

REMARKS: ☐ Urgent ☐ For your review ☐ Reply ASAP ☐ Comment

Acknowledgment. Breast cancer tissue
was obtained from the Breast Cancer
Resource of the Department of Pathology,
N.Y.U. Medical Center, Dr Helen
Feiner, Director. The Resource is funded
by The Department of the Army, Grant
DAMD 17-94-J-4177



560 First Avenue, New York, N.Y. 10016
Cable Address: NYUMEDIC

Department of Pathology

(212) 263-

8/12/96

Dr. Charles Carmeci
Dept of Surgery
Stanford University
1201 Welch Rd
Stanford, CA 94305

Dear Dr. Caremeci:

This is to confirm that on August 5, 1996 we shipped you 18 frozen breast tissue specimens, as follows:

- 8 estrogen receptor positive carcinomas
- 8 estrogen receptor negative carcinomas
- 2 non tumor breast tissue

Please let us know the state in which the specimens were received, a brief statement of the intended use, and how well the material served your purposes.

Many thanks in advance for this important feedback.

Yours sincerely,

A handwritten signature in cursive script, appearing to read "Helen Feiner".

Helen Feiner, M.D.
Director, Anatomic Pathology
Director, Breast Cancer Resource
PH (212) 263-8826 FAX (212) 263-7916

cc: Rita Demopoulos, M.D.

**STANFORD UNIVERSITY SCHOOL OF MEDICINE**

DEPARTMENT OF SURGERY
MEDICAL SCHOOL OFFICE BLDG. (MSOB),
SUITE X300
STANFORD, CALIFORNIA

Ph: ~~415/725-7280~~ FAX No: ~~415/725-3918~~

725-8762

TO: Helen Feiner

PH. NO: (212) 263 - 5470

FAX NO: (212) 263 - 7916

NO. OF PGS: 3 (Including this page)

FROM: Devon Thompson

PH. NO: (415) 498 - 5510

COMMENTS:

I f you have an email address I
could forward you the list of tumours
so you would have it on your computer

STANFORD UNIVERSITY
Department of Surgery

*1201 Welch Road
MSLS Building, Room P228
Stanford, CA 94305-5486
phone: (415) 498-5510 or (415) 725-1671
fax: (415) 725-8762*

*Devon A. Thompson, Ph.D.
Postoral Doctoral Fellow
devont@leland.stanford.edu*

Helen Feiner, M.D.
Department of Surgical Pathology
N.Y.U. Medical Center
560 First Avenue,
New York, NY 10016

July 9th 1997

Dear Dr. Feiner,

I have been working in collaboration with Charles Carmeci, M.D., with whom you have had previous discussions. We have received 31 breast tumour and 4 normal breast samples from your Breast Cancer Tissue Bank. This source has been invaluable to us. We have used these samples to extract RNA and then perform RT-PCR to detect several different genes. At this juncture it would be extremely useful if we could obtain any information you have in your files pertaining to the specific tumours that you have provided to us. Information such as, histological grade, the method(s) used to establish the estrogen receptor phenotype and quantitative values for the ER levels determined. Following I have listed our ID number and your ID number for each of the tumours that we have received.

I will be away on vacation from July 12th until July 26th. You can contact me by e-mail devont@leland.stanford.edu, by phone (415) 498-5510, or fax (415) 725-8762 after this date. Thank you for your help with this matter.

Sincerely,



Devon A. Thompson, Ph.D.
devont@leland.stanford.edu

Stanford ID	NYU Tumour ID	Current Information
1	s95 10787	ER + tumour
2	s95 11102	ER + tumour
3	s95 11319	ER + tumour
4	s95 12182	ER + tumour
5	s95 17621	ER + tumour
6	s96 2526	ER + tumour
7	s95 8934	ER + tumour
8	s95 12034	normal breast
9	s95 14255	normal breast
10	s95 14788	ER - tumour
11	s95 21162	ER - tumour
12	s96 1236	ER - tumour
13	s96 2129	ER - tumour
14	s96 2423	ER - tumour
15	s95 10239	ER - tumour
16	s95 12254	ER + tumour
17	s96 12828	ER - tumour
19	s97 4250	ER - tumour
20	s97 4288	ER + tumour
21	s96 20823	ER + tumour
22	s97 2396	ER + tumour
23	s97 4379	ER + tumour
24	s97 3778	ER + tumour
25	s96 20371	ER + tumour
26	s97 4926	ER + tumour
27	s97 596	ER + tumour
28	s96 19122	ER - tumour
29	s97 528	ER - tumour
30	s96 12363	ER - tumour
31	s96 14116	ER - tumour
32	s96 20358	ER - tumour
33	s96 15075	ER - tumour
34	s97 1647	ER - tumour
35	s97 1792	normal breast
		normal breast

cc:Mail for: Helen Feiner

Subject: NYU breast cancer samples.

From: Helen Feiner 7/24/97 12:50 PM

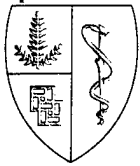
To: devont@leland.stanford.edu at PMDF

Dear Dr Thmpson,

I have sent you, by mail, two reports from each of the patients listed in your communication of July 9th. One is the surgical pathology report from which you can derive a histologic grade. The most common grading system utilizes architectural grade + nuclear grade + mitotic rate. The second report is from our molecular pathology lab from which you can obtain the estrogen receptor quantitative values. Let me know if you need help with any of these data.

Method used to obtain ER phenotype: Indirect immunoperoxidase technique. Estrogen receptor antibody is obtained from AMAC (clone ER1D5, Westbrook, ME) and Novo Castra (clone 6F11, distributed by Vector, Burlingame, CA). Secondary antibody is horse anti mouse IgG. A standard avidin-biotin-peroxidase technique is used on formalin fixed, paraffin embedded tissue sections. Antibody expression is evaluated in 10 40x fields in a CAS Image Analyzer. Result is expressed as percent positive nuclear area.

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Helen Feiner, M.D.
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August 25 1998

Dear Dr. Feiner,

Thank you for the information pertaining to the breakdown of race status, with regard to patients from whom the breast tumour specimens are obtained. Enclosed are re-prints from some papers in which we have used the tumour specimens that you provided. These frozen tumours have been invaluable to us in extrapolating our findings in breast cancer cell lines to breast tumour biology. We hope to continue using the Breast Cancer Resource of the Department of Pathology, New York University Medical Center, to obtain breast cancer samples.

Sincerely,

A handwritten signature in cursive script, reading "Devon A. Thompson".

Devon A. Thompson, Ph.D.

PUBLICATIONS THAT ACKNOWLEDGE THE RESOURCE :

Eur. J. Biochem. 252, 169–177 (1998)
© FEBS 1998

Characterization of a gene that is inversely correlated with estrogen receptor expression (ICERE-1) in breast carcinomas

Devon A. THOMPSON and Ronald J. WEIGEL
Department of Surgery, Stanford University, Stanford CA, USA

(Received 22 September/10 December 1997) – EJB 97 1350/1

1116–1123 *Nucleic Acids Research*, 1998, Vol. 26, No. 4

© 1998 Oxford University Press

Differential screening and suppression subtractive hybridization identified genes differentially expressed in an estrogen receptor-positive breast carcinoma cell line

Wayne W. Kuang, Devon A. Thompson, Renee V. Hoch and Ronald J. Weigel*

Department of Surgery, Stanford University, Stanford, CA 94305, USA

Received June 10, 1997; Revised and Accepted December 18, 1997

DDBJ/EMBL/GenBank accession no. AF007170

GENOMICS 45, 607–617 (1997)
ARTICLE NO. GE974972

Identification of a Gene (GPR30) with Homology to the G-Protein-Coupled Receptor Superfamily Associated with Estrogen Receptor Expression in Breast Cancer

Charles Carmeci,* Devon A. Thompson,* Huijun Z. Ring,†
Uta Francke,†† and Ronald J. Weigel*,¹

*Department of Surgery, †Department of Genetics, and ††Howard Hughes Medical Institute,
Stanford University, Stanford, California 94305

Received April 4, 1997; Accepted August 11, 1997



Request for Tissue/Cells
NYU Breast Cancer Resource
Director: Helen D. Feiner, M.D.
560 First Ave. NY, NY 10016
Tel (212) 263-8826
Fax (212) 263-7916

Name: Dr. KEN TAKASHITA

Title: ASSISTANT PROFESSOR

Address: NYU MEDICAL CENTER
DEPT. OF HEMATOLOGY

Phone: (212) 263-5465

Fax: (212) 263-8444

e-mail: --

Grant Support: YES

Material Requested: FROZEN SECTIONS OF
METASTATIC BREAST CANCER.

Date Shipped: 8-15-97

Date Received: SAME DAY

State of Specimen on receipt: GOOD

Brief Summary of intended use:

(Use additional page if necessary)

TO PERFORM IN SITU HYBRIDIZATION AND
IMMUNOHISTOCHEMISTRY, IN ORDER TO DETERMINE
WHETHER THE DECREASED EXPRESSION OF RAR-ALPHA
RETINOIC ACID RECEPTOR EXPRESSION SEEN IN
METASTATIC BREAST CA. IS DUE TO A TRANSCRIPTIONAL
DEFECT OR A TRANSLATIONAL DEFECT.

NYU
Medical
Center

Hematology Division, Department of Medicine
New York University Medical Center
550 First Avenue, New York, N.Y. 10016 U.S.A.

e-mail takeshtk@is.nyu.edu
Tel 1-212-263-5465, Fax 1-212-263-8444

12/9/97
Re Dr. Takeshita & confirmed
by H.F. on slide review
Good signal in IHC
r ISH

August 15, 1997

Dr. Helen Feiner
Department of Pathology
Breast Cancer Archives

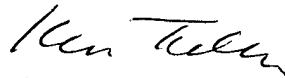
Dear Dr. Feiner:

I am writing to notify you that we have requested and received from Yara Delgado of the breast tumor registry frozen sections of lymph nodes containing known breast cancer metastasis from 7 different patients. We received 6 slides for each patient.

These sections will be used to perform in situ hybridization and immunohistochemistry. The objective of this experiment is to determine whether the decreased expression of RAR-alpha retinoic acid receptor expression seen in metastatic breast cancer is due to a transcriptional defect or a translational defect.

We are grateful for your assistance in our studies. Please contact me if you have any questions.

Sincerely yours,



Ken Takeshita, M.D.
Assistant Professor of Medicine

Feiner, Helen, D.
DAMDM17-94-J-4177

best available copy

APPENDIX 2

NYU BREAST CANCER RESOURCE FOR
RESEARCH AND BANKING

Phone: (212) 263 8826-8079

Fax #: (212) 263 7916

RECORD OF EVALUATION OF BANKED MATERIAL:

Type of Specimen: Imprint ✓ Frozen Tissue _____

Date of evaluation:

9/4/97

Duration in freezer:

2 years

Type of evaluation:

IMMUNO HISTOCHEMISTRY

Results:

Excellent

Entered by:

HELEN FEINER MD

Signature and date:

H Feiner

9/13/97.

IHC#

IMMUNOHISTOCHEMISTRY

* Place completed form

in Tisch-379 (ext 8922)

RESIDENT/ ATTENDING Dr. FennelPATIENT Anonymous SURG PATH#: 98-45-16437 Block *DATE 5/12/98 SITE Breast SPECIMEN: biopsy/ major ImpmtDIAGNOSTIC ISSUE None / QC material

ANTIBODIES : CIRCLE (if limited tissue, number antibodies according to priority, and request "numbered PLL" slides under special requests)

LEU- M1	Calretinin	CK19	GFAP	B72.3	Adenovirus
Muscle Specific	<u>CAM 5.2 (CK)</u>	NSE	*HCG	PSA	*HBsAg
ACTIN					
DESMIN	AE1/AE3	CHRO	PLAP	PAP	*HBcAg
SMA	EMA	*SYN	*AFP	FVIII	CMV
VIMENTIN	34BE12	*CALCITON	pCEA	CD34	*HSV
*S-100	CK7	THYRO	mCEA	CD 68	ER/PR
HMB45	CK20	*MYOGLOBIN	<u>LCA</u>	BerEP4	Brst-2

PLL SLIDES REQUESTED (circle # if ordered on gross sheet)SPECIAL REQUESTS:(CIRCLE): RUSH / USE H&E SECTION / RCVD STAINED SIGNED OUT TURNAROUND SPECIAL PROCESSING IHC INTERPRETATION: NEGATIVE CONTROL - ⊖ POSITIVE CONTROLS - ⊕

Cam 5.2 - 3+

LCA - 2-3+

remedial step(s): result- Conclusion-

ABBREVIATIONS AND CLONE # * = POLYCLONAL ANTIBODY

SMA= SMOOTH MUSCLE ACTIN (1A4), MUSCLE SPECIFIC ACTIN (HHF-35), AE1/AE3 & CAM 5.2= LOW MOL WT KERATINS, 34BE12= HIGH MOL WT KERATIN, CALC= CALCITONIN, THYRO= THYROGLOBULIN (Tg6), PSAP= PR ACID PHOS (PASE/4LT), PSA= PROSTATE SPECIFIC ANTIGEN (EA-PR8), FVIII=FACTOR VIII (F8/86), SYN= SYNAP CYTOMEGALOVIRUS (DDG9, CCH2), DESMIN (D33), VIMENTIN (V9), LCA= LYMPHOCYTE COMMON ANTIGEN (2B11), CEA= (A5B7), EMA= (E29), AFP= ALPHAFETOPROTEIN (M1A1301), HSV= HERPES. PLAP= PLACENTAL A PHOSPHATASE (8B6)

HCG= HUMAN CHORIONIC GONADOTROPHIN, HBsAg =HEPATITIS B SURFACE Ag, HBcAg=HEPATITIS B CORE Ag CD34=(QBEND10.), CD68=(KPI), CK7= OV-TL 12/30, CK20= 1T-KS20.8

Histology: Date/time submitted date/time cut



NYU BREAST CANCER RESOURCE FOR RESEARCH AND BANKING

Phone: (212) 263 8826-8079

Fax #: (212) 263 7916

RECORD OF EVALUATION OF BANKED MATERIAL:

Type of Specimen:

Aspirated Cells _____
Imprint _____

Frozen Tissue ✓

Date of evaluation:

September/ 97

Duration in freezer:

2 YEARS

Type of evaluation:

IMMUNOHISTOCHEMISTRY: MIB-1
p21

Results:

SATISFACTORY TO GOOD

Entered by:

Dr. H. FEINER

Signature and date:

MF 9/12/97

IHC#

IMMUNOHISTOCHEMISTRY* Place completed formin Tisch-379 (ext 8922)RESIDENT/ ATTENDING H Fine

95-11102

PATIENT _____

SURG PATH#: 98-Block DATE 9/5/97 (*only single block/case will be stained - except by special request)SITE Breast SPECIMEN: biopsy/ majorDIAGNOSTIC ISSUE Q A material - (FS)**ANTIBODIES** : CIRCLE (if limited tissue, number antibodies according to priority, and request "numbered PLL" slides under special requests)

LEU- M1	Calretinin	CK19	GFAP	B72.3	Adenovirus
Muscle Specific	CAM 5.2 (CK)	NSE	*HCG	PSA	*HBsAg
ACTIN	AE1/AE3	CHRO	PLAP	PAP	*HBcAg
DESMIN	EMA	*SYN	*AFP	FVIII	CMV
SMA	34BE12	*CALCITON	pCEA	CD34	*HSV
VIMENTIN	<u>CK7</u>	THYRO	mCEA	CD 68	ER/PR
*S-100	CK20	*MYOGLOBIN	LCA	BerEP4	Brst-2
HMB45					

PLL SLIDES REQUESTED _____ (circle # if ordered on gross sheet)

SPECIAL REQUESTS:(CIRCLE): RUSH / USE H&E SECTION / _____

RCVD _____ STAINED _____ SIGNED OUT _____ TURNAROUND _____

SPECIAL PROCESSING _____

IHC INTERPRETATION: NEGATIVE CONTROL- POSITIVE CONTROLS- (+)

P 21 - 2 +
CK 7 - 2 +

remedial step(s):

result-

Conclusion- H Fine

ABBREVIATIONS AND CLONE #

* = POLYCLONAL ANTIBODY

SMA= SMOOTH MUSCLE ACTIN (1A4), MUSCLE SPECIFIC ACTIN (HHF-35), AE1/AE3 & CAM 5.2= LOW MOL WT
 KERATINS, 34BE12= HIGH MOL WT KERATIN, CALC= CALCITONIN, THYRO= THYROGLOBULIN (Tg6), PSAP= PR
 ACID PHOS (PASE/4LT), PSA= PROSTATE SPECIFIC ANTIGEN (EA-PR8), FVIII=FACTOR VIII (F8/86), SYN= SYNAP
 CHRO= CHROMOGRANIN (tk2H10), CMV= CYTOMEGALOVIRUS (DDG9, CCH2), DESMIN (D33), VIMENTIN (V9), LCA= LYMPHOCYTE COMMON ANTIGEN
 2B11), CEA= (A5B7), EMA= (E29), AFP= ALPHAFETOPROTEIN (M1A1301), HSV= HERPES, PLAP= PLACENTAL A
 PHOSPHATASE (8B6)
 HCG= HUMAN CHORIONIC GONADOTROPHIN, HBsAg =HEPATITIS B SURFACE Ag, HBcAg=HEPATITIS B CORE Ag
 CD34=(QBEND10.), CD68=(KPI), CK7= OV-TL 12/30, CK20= IT-Ks20.8

Histology:

Date/time submitted

date/time cut

NYU BREAST CANCER RESOURCE FOR
RESEARCH AND BANKING

Phone: (212) 263 8826-8079

Fax #: (212) 263 7916

RECORD OF EVALUATION OF BANKED MATERIAL:

Type of Specimen: Imprint ✓ Frozen Tissue

Date of evaluation: 9/9/98

Duration in freezer: 2 years

Type of evaluation: FISH

Results: Attached

Entered by: J. FEINER

Signature and date: [Signature] 9/18/98



NEW YORK UNIVERSITY MEDICAL CENTER CYTOGENETICS LABORATORY

New Bellevue Hospital
Dept. of Pathology/Cytogenetics Lab
Room 4 North 20
27th Street & First Avenue, New York, NY 10016

(212) 263-6454

(212) 562-3496

Fax: (212) 263-7930

PATIENT S96-20371 CASE # Research-S9620371
REFERRAL Dr. H. Feiner DATE COMPLETED 9/09/98
HOSPITAL NYU-Research DATE COLLECTED unknown
DATE RECEIVED 9/03/98

MOLECULAR CYTOGENETIC ANALYSIS

SPECIMEN TYPE Tissue Imprints CHROMATID BREAKS _____
QUALITY OF PREPARATION adequate CHROMOSOME BREAKS _____
NO. OF CELLS EXAMINED 50+ ANEUPLOID CELLS _____

INTERPRETATION:

Slides were received from air-dried material described as imprints/scrapes from breast tissue.

Interphase molecular cytogenetic analysis was performed using fluorescent in situ hybridization (FISH) with investigational DNA probes specific for the centromeric region of the X chromosome (Vysis CEP X-alpha probe set). Random sections of the slide were examined by two independent readers. Adequate signal for analysis was seen over the majority of the hybridization area. Results indicated over 85% of cells contained two signals for the X chromosome consistent with two copies of the X. No evidence was seen of X chromosome aneuploidy.

MOLECULAR CYTOGENETIC DIAGNOSIS: nuc ish Xcen(DXZ1x2)

Mary Ann Perle, Ph.D.
Director, Cytogenetics
Laboratory

Note: Since this is an in vitro test, accuracy may be limited by technical or cultural artefacts.

Feiner, Helen, D.
DAMDM17-94-J-4177

APPENDIX 3

BREAST CANCER PILOT PROJECTS AWARDED

1995 - 1998

1995 GRANT YEAR

Pamela Cowin, Ph.D.
Assistant Professor
Cell Biology

"The Role of Plakoglobin in Breast Cancer"
(\$30,000)

Xiao-Hong Sun, Ph.D.
Assistant Professor
Cell Biology

"The Role of ID Proteins in Breast Cancer"
(\$28,450)

Mary Ann Perle, Ph.D.
Assistant Professor
Pathology

"Chromosomes 7, 18, 20 and X in Mammogram
Detected Atypical Ductal Hyperplasia and
Ductal Carcinoma in situ"
(\$8,950)

1996 GRANT YEAR

Sandra Reynolds, Ph.D.
Res. Assistant Professor
Dermatology

"Peptide Epitopes Recognized by CD8+ T Cells
in Patients with Breast Cancer"
(\$10,000)

Herbert Samuels, M.D.
Professor
Medicine

"Retinoid-Regulated Genes and Breast Cancer"
(\$25,000)

Jan Sap, Ph.D.
Assistant Professor
Pharmacology

"Receptor Protein Tyrosine Phosphatases and
Breast Cancer"
(\$20,000)

Kenichi Takeshita, M.D.
Assistant Professor
Medicine

"9-cis Retinoic Acid and Retinoid X Receptor RXR
in Breast Cancer"
(\$20,000)

Stephen Tomlinson, Ph.D.
Assistant Professor
Pathology

"The Role of Complement Inhibitors in Tumorigenicity"
(\$10,000)

Stanislav Vukmanovic, MD, PhD
Assistant Professor
Pathology

"Effector Function of Vaccine Induced CD8+
Cells"
(\$10,000)

1997 GRANT YEAR

Harry Ostrer, M.D. (P.I.)
Professor
Pediatrics
Ruth Oratz, M.D. (Co-P.I.)
Assistant Professor
Medicine

"Genetic Susceptibility to Breast Cancer"
(\$15,000)

W. Fraser Symmans, M.D. (P.I.)
Assistant Professor
Pathology
Matthew Volm, M.D. (Co-P.I.)
Instructor
Medicine

"A Response Biomarker for Paclitaxel Chemotherapy
in Patients with Breast Cancer"
(\$29,875)

Carolyn Wasserheit, M.D. (P.I.)
Assistant Professor
Medicine
Kenichi Takeshita, M.D. (Co-P.I.)
Assistant Professor
Medicine

"Biological Correlates of 9-Cis Retinoic Acid and
Tamoxifen"
(\$15,000)

1998 GRANT YEAR

Ruben Abagyan, Ph.D.
Associate Professor
Biochemistry

"Toward A New Chemotherapy for Breast Cancer:
Rational Design of A Retinoid X Receptor-Selective
Agonist"
(\$29,968)

Alan Frey, Ph.D.
Assistant Professor
Cell Biology

"Translational Arrest of IL-2 mRNA in Human Breast
Cancer Tumor Infiltrating Lymphocytes"
(\$30,000)

Giorgio Inghirami, M.D.
Associate Professor
Pathology

"Molecular Characterization of BRCA1"
(\$30,000)

Carole Oddoux, Ph.D.
Assistant Professor
Pediatrics

"Heritable Susceptibility to Invasive and Non-Invasive Breast
Cancer"
(\$15,000)